

Supplemental Figure 1. Effects of IL-21 treatment on memory CD4⁺ T-cell subsets. Longitudinal assessment of the fractions of CD4⁺ T-cells with a central memory (CD28⁺CD95⁺CCR7⁺; **A,C,E**) or effector memory (CD28⁺CD95⁺CCR7⁻; **B,D,F**) phenotype in peripheral blood (PB; **A,B**), colorectal (RB; **C,D**), and lymph node (LN; **E,F**). IL-21 treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean ± SEM.



Supplemental Figure 2. Effects of IL-21 treatment on CD8⁺ T-cell levels. Longitudinal assessment of the frequencies (A) and absolute counts (cells per μ l of blood; B) of peripheral blood (PB) CD8⁺ T-cells. Frequencies of CD8⁺ T-cells were also determined in colorectal (RB; C), and lymph node (LN; D) biopsies. IL-21-treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean ± SEM.



Supplemental Figure 3. Effects of IL-21 treatment on the frequencies of CD8⁺ T-cells expressing perforin, granzyme B, and T-bet. Longitudinal assessment of the frequencies of peripheral blood (A-C) and LN (D-F) CD8⁺ T-cells expressing perforin (A,D), granzyme B (B,E), or T-bet (C,F). (g) Representative staining for T-bet by perforin and T-bet by granzyme B in blood CD3⁺CD8⁺ T-cells from a representative IL-21-treated RM at d105 p.i. IL-21-treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean ± SEM.



Supplemental Figure 4. Effects of IL-21 treatment on blood and LN B-cell levels. Longitudinal assessment of the percentages of blood (A-E) and LN (G-K) $CD3^{-}CD20^{+}$ B-cells (A,G) as well as their naïve (CD21+CD27-; B,H), tissue memory (CD21-CD27-;

C,I) activated memory (CD21-CD27+; **D,J**), and resting memory (CD21+CD27+; **E-K**) subsets. (**F,L**) Representative staining for CD21 by CD27 in CD3⁻CD20⁺ B-cells from a representative IL-21-treated RM at d105 p.i. IL-21 treated animals (n=7) are depicted as (●), controls (n=8) as (■). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean ± SEM



Supplemental Figure 5. Effects of IL-21 treatment on the localization of B-cells in the LN. IHC analyses on LN biopsies showed that early on-ART IL-21-treated animals have higher levels of B-cells in the medulla when compared to controls (P=0.0238; n=5). Random $20 \times$ and $200 \times$ images of LN were taken and the percent area staining for CD20 was determined in IL-21-treated (n=5) (\bullet) and control (n=5) (\bullet) RMs (left). The right panels show a representative CD20 staining ($200 \times$) in one IL-21-treated (top panel) and one control (bottom panel) RM on-ART (d105 p.i.). Statistical analysis was performed by Mann-Whitney t-test.



Supplemental Figure 6. Higher levels of intestinal CD4⁺ T-cells co-expressing IL-17 and IL-22 in IL-21-supplemented RMs. Longitudinal assessment of the frequency of intestinal CD4⁺ T cells producing IL-17 and IL-22. The levels of CD4⁺IL-17⁺IL-22⁺ Tcells were significantly higher in the rectum of IL-21-treated (n=7) RMs (\bigcirc) as compared to controls (n=8) (\blacksquare) at d85 and 135 p.i. (P=0.0209 and P=0.0281, respectively). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean ± SEM. Repeated-measures analyses were performed with a means model (SAS Proc Mixed v9) to generate statistical outcomes.



Supplemental Figure 7. Effects of IL-21 treatment on T-cell activation in the LN. Longitudinal assessment of the frequencies of LN-derived memory $CD4^+(A)$ and $CD8^+(B)$ T-cells expressing the activation markers $HLA-DR^+CD38^+$. IL-21-treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare). Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean \pm SEM. Repeated-measures analyses were performed with a means model (SAS Proc Mixed v9) to generate statistical outcomes.



Supplemental Figure 8. Reduced levels of intestinal T-cell proliferation and soluble markers of inflammation in IL-21-treated RMs. (A,B) Longitudinal assessment of the frequencies of intestinal $CD4^+$ (A) and $CD8^+$ (B) T-cells expressing the proliferation marker Ki-67 in IL-21-treated (n=7) (\bigcirc) and control (n=8) (\square) RMs. Fractions of proliferating CD4⁺ and CD8⁺ T-cells were significantly lower in the rectum of IL-21treated animals at d85 and 256 p.i. Shaded area represents time of ART treatment and orange arrows represent IL-21 administrations. Averaged data are presented as mean \pm SEM. (C,D) Plasma levels of IP-10 (C) and CRP (D) were compared between IL-21treated and control RMs at pre-ART and different experimental points on-ART. Although similar to those found in controls at pre-ART, IL-21-treated RMs showed significantly lower levels of IP-10 at d143 and d200 on-ART (P=0.0401 for both experimental points) and of CRP at d75 on-ART (P=0.0323). Averaged data are presented as mean \pm SEM. Repeated-measures analyses were performed with a means model (SAS Proc Mixed v9) to generate statistical outcomes.



Supplemental Figure 9. GSEA analysis of enriched pathways in IL-21-treated versus control RMs. Representation of the rank in gene list of JAK/STAT pathway at d50 (A) and d200 (B) on-ART, as well as for interferon-stimulated genes (ISGs) at d50 (C) and d200 (D) on-ART. Leading genes with higher enrichment score are depicted as (•). Statistical analyses were performed with GSEA tool.

Rectal Biopsy



Supplemental Figure 10. Reduced levels of $CD4^+ PD-1^+ T$ -cells in the rectum of IL-21-treated RMs. Fractions of intestinal $CD4^+ T$ -cells expressing PD-1 were longitudinally determined throughout the study. Although similar (or even higher) to those found in controls at pre-ART, the fractions of $CD4^+ PD-1^+ T$ -cells at d200 on-ART were significantly lower (P=0.0232) in IL-21 supplemented RMs. IL-21-treated animals (n=7) are depicted as (\bullet), controls (n=8) as (\blacksquare). Averaged data are presented as mean \pm SEM. Repeated-measures analyses were performed with a means model (SAS Proc Mixed v9) to generate statistical outcomes.



Supplemental Figure 11. Markers of mucosal immunity and inflammation correlate with measures of SIV persistence. SIV-DNA content in purified blood CD4⁺ T-cells at the latest time point on-ART (d256 p.i.) correlates negatively with the levels of intestinal Th17 (A) and Th22 cells (C) at pre-ART, negatively with the levels of intestinal Th17 cells on-ART (B), and positively with the levels of intestinal CD4⁺Ki-67⁺ T-cells on-ART (D). At the latest time point on-ART, the levels of activated (HLA-DR+CD38+) circulating CD4⁺ (E) and CD8⁺ (F) T-cells positively correlate with residual plasma viremia. Plasma levels of IP-10 (G) and CRP (H) on-ART positively correlate with SIV-DNA content in intestinal tissues and residual plasma viremia, respectively. IL-21-treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare).



Supplemental Figure 12. Pre-ART plasma viremia strongly correlates with viral rebound post-ART interruption. Plasma viremia at pre-ART (d58 p.i.) positively correlates with plasma viremia at days 90 (A) and 240 (B) post-ART interruption. IL-21-treated animals (n=7) are depicted as (\bigcirc), controls (n=8) as (\blacksquare).



С

ratio #CD4/CD8 T cells PB 2

3

1

0

days off-ART

-14



P=0.0541

28

F60

P=0.0401

180

240



Supplemental Figure 13. Reduced lymph node T-cell activation and higher blood CD4⁺ to CD8⁺ T-cell ratios in IL-21-treated RMs following ART interruption. (A,B) Fraction of lymph node derived $CD4^+$ (A) and $CD8^+$ (B) T-cells expressing HLA-DR and CD38 at necropsy (d250 off-ART). T cell activation levels were lower in IL-21-treated (n=5-7) RMs (\bigcirc) as compared to controls (n=6-8) (\blacksquare), with the differences reaching statistical significance for CD4+ T-cells, (P=0.0260). (C) The ratios between the absolute counts of blood CD4⁺ and CD8⁺ T-cells were longitudinally compared between IL-21treated and control RMs. CD4⁺ to CD8⁺ T-cell ratios were higher in IL-21-treated RMs. with the differences reaching statistical significance at d180 off-ART (P=0.0401). Averaged data are presented as mean \pm SEM. Repeated-measures analyses were performed with a means model (SAS Proc Mixed v9) to generate statistical outcomes.

Supplemental Table	able 1
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Animal	Age	Sex	#CD4	VL	MamuA01	IL-21	Diarrhea	Anemia	Weight
ID			preART	preART	genotype	Тх	Rx	Rx	loss*
RLm12	57 ms	F	540	1.07E+05	+	у	n	n	у
ROe12	65 ms	F	707	1.73E+03	-	у	n	У	n
RJp11	81 ms	F	709	1.99E+04	+	у	n	n	у
RGv10	102 ms	F	379	6.94E+05	-	у	n	у	у
RVt10	98 ms	F	366	6.83E+05	+	у	n	n	n
RTb12	71 ms	F	364	1.66E+06	-	у	n	У	у
$ROc10^{\Psi}$	114 ms	F	209	1.27E+06	+	у	n		
RHa10	114 ms	F	618	2.31E+04	-	у	n	У	n
RBt12	57 ms	F	678	6.56E+03	+	n	n	У	у
RGe12	67 ms	F	594	3.20E+06	-	n	n	n	n
RCb12	70 ms	F	942	3.38E+03	+	n	n	n	у
RJw12	54 ms	F	463	4.54E+05	-	n	n	n	у
RKg11	90 ms	F	1108	2.12E+04	+	n	n	n	у
RPu12	57 ms	F	280	2.47E+06	-	n	n	У	у
RKd12	68 ms	F	452	1.53E+05	-	n	n	У	у
RPy8	120 ms	F	374	3.50E+05	+	n	n	у	n

Animals included in the study. ^{Ψ} ROc10 died at d140 p.i; **ms** (months) at time of infection; **F** (Female); **#CD4 pre-ART** in blood (cells/µL); Viral Load **pre-ART** (day 58 p.i.), SIV-RNA copies/mL of plasma; + (positive) and – (negative); **y** (yes) and **n** (no); * weight loss above 10% of the pre-infection weight. All RMs were treated with ART.

Supplemental Table 2

Immunologic and virologic markers	Day p.i.	Tissue	Related gene	Correlation r values
			PRDM1	-0.514763
Memory %CD4	Day 105	Colorectal biopsy	SOCS1	-0.700644
HLA-DR+CD38+			IL22	-0.602942
			SOCS1	-0.536038
Memory %CD4	D 056		MUC4	-0.601547
HLA-DR+CD38+	Day 256	Colorectal biopsy	TLR7	0.618818
			IL17F	-0.597841
Memory %CD8	D. 105	Calana (all binar	PRDM1	-0.520453
HLA-DR+CD38+	Day 105	Colorectal blopsy	SOCS1	-0.587045
Memory %CD8	Day 256	Coloractal biongy	TLR7	0.743636
HLA-DR+CD38+	Day 250	Colorectal blopsy	IL17F	-0.72236
%CD4 Ki-67+	Day 105	Colorectal biopsy	PRDM1	-0.551366
/0CD4 KI-0/ +	Day 105	Colorectar biopsy	IL23R	-0.595036
%CD4 Ki-67+	Day 256	Colorectal biopsy	AHR	-0.527883
			SOCS3	-0.528842
%CD8 Ki-67+	Day 105	Colorectal biopsy	PTGS2	-0.517683
	5	1 5	S100A8	-0.497802
			AHR	-0.590169
	D 056		CAMP	-0.518008
%CD8 K1-6/+	Day 256	Colorectal biopsy	TREM1	-0.575188
			CCR2	-0.575164
			SOCS3	-0.565448
LP %PMN_MPO	Day 105	Colorectal biopsy	PTGS2	-0.640115
			S100A8	-0.568288
			SOCS1	0.690619
%CD4 IL-17+	Day 105	Colorectal biopsy	TNF	-0.57258
			PRDM1	0.536043
			IL23R	0.727713
			IL6	-0.717971
%CD4 IL-22+	Day 256	Colorectal biopsy	DEFA1	0.654452
			RGS18	0.554019
~ !!			IL4	0.720438
Cell-associated SIV-RNA	Day 105	Colorectal biopsy	TREM1	-0.555096
Memory %CD4	Day 105	Blood	SOCS1	-0.545821
			Π 10	0.541000
			ILIU FOVD2	0.341909
%CD4 Ki-67+	Day 105	Blood	TOAT 5 TNF	0.561581
			ICOS	0.578906
%CD8 Ki-67+	Day 105	Blood	TNF	0.599172
/0CD0 KI-0/+	Day 105	Diood	SOCS3	-0 593307
			FOXP3	0.518763
Cell-associated			BCL6	-0.67013
SIV-DNA in	Day 105	Blood	AHR	-0.538248
CD4+			S100A8	-0.6728
- '			TREM1	-0.576074
			CCR2	-0.520335
Memory %CD4	Dars 105	T	SOCS1	-0.543691
HLA-DR+CD38+	Day 105	Lymph node	VNN2	-0.521476

Memory %CD8	Day 105	Lymph node	SOCS1	-0.79014
ILA-DK+CD30+				-0.001048
%CD4 Ki-67+	Day 105	Lymph node	АПК	-0.399083
	-	v 1	CCR2	-0.533355
0/CD4V; 67+	Day 256	Lymph nodo	EOMES	-0.711951
%CD4 KI-0/+	Day 250	Lymph node	IRF4	-0.664508
0/CD9 V; 67+	Day 105	Lymph node	AHR	-0.576554
70CD0 KI-07+			CCR2	-0.569371
%CD8 Ki-67+	Day 256	Lymph node	EOMES	-0.747841
Residual viral	D. 105	Dlama	IL21R	-0.578903
SIV-RNA	Day 105	Plasma		
Residual viral	D 056	Diama	CLU	-0.645965
SIV-RNA	SIV-RNA Day 256		CCR2	-0.591526

Immunologic and virologic markers associated with RNA-Seq transcripts. Day p.i; data of the experimental point used; **Tissue**; data of the anatomical compartment used; **Related gene**; RNA transcript correlated with the immunological and virologic markers; **Correlation r values**; r value of each correlation. P-value (<0.05) for all the correlations.

Supplemental Table 3

Immunologic and Virologic Markers	Day p.i.	Immunologic and Virologic Correlates	Correlation r values	P values
SIV-DNA copies in CD4 ⁺ T cells (PBMC)	256 (200 on-	Log ₁₀ VL d58 only IL-21 Tr.	0.6598	0.1068
	ART)	Log ₁₀ VL d58 only Controls	0.7022	0.0612
SIV-DNA copies	256 (200 on-	Log ₁₀ VL d58 only IL-21 Tr.	0.7500	0.0663
(RB)	ART)	Log ₁₀ VL d58 only Controls	0.0931	0.8263
Log ₁₀ plasma Viral Load	58 (pre-ART)	%CD8 ⁺ DR ⁺ CD38 ⁺ d105 (RB) %CD8 ⁺ DR ⁺ CD38 ⁺ d256 (RB) Th17 d58 (RB) Th17 d105 (RB) Th22 d58 (RB) %CD4 Ki-67 ⁺ d84 (RB) IP-10 pg/mL d203 (plasma) CRP ug/mL d135 (plasma) Log ₁₀ VL d90 off-ART Log ₁₀ VL d240 off-ART	0.2437 0.2643 -0.6988 -0.2739 -0.4500 0.4392 -0.2119 0.3204 0.9048 0.8179	$\begin{array}{c} 0.3815\\ 0.3402\\ 0.0047\\ 0.3233\\ 0.0944\\ 0.1014\\ 0.4483\\ 0.2443\\ 0.0001\\ 0.0002\\ \end{array}$

Immunologic and virologic markers associated with pre-ART plasma viremia. Day p.i.; data of the experimental point used.

Supplemental Table 4

Animal ID	VL d135 p.i.	PB #CD4 d135 p.i.	PB %CD8 DR ⁺ 38 ⁺ d135 p.i.	RB %CD4 d135 p.i.	RB %Th17 d135 p.i.	RB %Th12 d135 p.i.	RB %CD8 DR ⁺ 38 ⁺ d135 p.i.
$ROc10^{\Psi}$	82	322	3.57	32.5	19.7	15.2	8.6
Av. IL-21 Tr.	30	683	4.6	21.4	13.3	7.2	7.8
Av. Controls	50	732	9.5	26.8	6.7	3.2	15.8

^{Ψ} ROc10 was euthanized at day 140 p.i. and the data reported were collected at necropsy; **Av.** (average) of IL-21 treated (excluding ROc10) and control groups at day 135 p.i. (closest experimental point to ROc10's necropsy at d140 p.i.); **VL** (viral load), limit of detection 60 copies/mL of plasma; **PB** (peripheral blood); **RB** (rectal biopsy).